Tuesday 28th October 2025

CURRICULUM VITAE – GAURAV CHATTERJEE

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Educational qualification:

Year	Degree Obtained	% marks	Institute studying in at the time
2005	Higher Secondary Examination **(HSC)**	87.3%	The Vivekananda Institution, Howrah, India
2011	Bachelor of Medicine; Bachelor of Surgery (**M.B.B.S**)	68.2%	RGKMC&H, Kolkata, India
2015	Doctor of Medicine (**M.D.**)- Pathology	70.6%	IPGMER & SSKM Hospital, Kolkata, India
2017	H.B.N.I fellowship in Haematopathology	79%	Tata Memorial Centre, Mumbai
2018	Master of Research in Translational Cancer Medicine	74%, **Distinction**	Kings College London, UK

Faculty Positions held:

- **Current:** Associate Professor, Hematopathology, Tata Memorial Centre, Mumbai (1st November 2022 – current)
- **Past:**
- Assistant Professor, Hematopathology, Tata Memorial Centre, Mumbai (25th June 2020 – 31st October 2022)
- Lecturer, Hematopathology, Tata Memorial Centre, Mumbai (1st October 2018 – 24th June 2020)

Research grants awarded:

1. Recipient of **DBT/Wellcome Trust India Alliance Intermediate Fellowship for Clinical and Public Health** (2023) for the project titled as **"Concurrent interrogation of genetic and epigenetic features in Diffuse large B-cell lymphoma and dynamics of recurrence/progression"** (Grant of 3,80,00,000 INR, ~460,000 USD).
2. I am part of a multi-institutional team (individual project PI) that were recently awarded (2025) the **ICMR Centre for Advanced Research for on Innovation in Diagnosis, Prognostication, and Monitoring of AML** (A total grant of 15,00,00,000 INR, ~1,750,000 USD).
3. Have been awarded **Institutional Research Grant** of 1,50,00,000 INR (~181,500 USD) for the project titled as **"Deciphering the molecular heterogeneity in Acute Leukemia of Ambiguous Lineage at a cellular level"** by **Lady Tata Memorial Trust**, 2022.

Selected awards:

Selected as research fellow from Tata Memorial Centre to pursue **Master of Research in Translational Cancer Medicine at King's College London**, **UK,** 2017-18

Professional experience and skills:

1. **Routine morphological assessment of all samples in hematopathology laboratory** including bone marrow aspirate, peripheral blood film and body fluid smears as well as cytochemistry (MPO, NSE)**.**

2. **Multicolor (10-16+ color) flow cytometry:**

a) I have been extensively involved in design, development, and routine reporting of **multi-color flowcytometry (13-18c) panels for all hematolymphoid neoplasms.** I have helped in setting up panels for acute leukemia, CLPD, lymphocyte screening (LST), plasma cell neoplasm and MDS compatible with 13-color CytoFlex (BC) and Dxflex (BC), 16-color Fortessa (BD), 18-color CytoFlex LX (BC) etc.

b) My work has significantly advanced the process of establishing **high-sensitivity measurable residual disease (MRD)** assays in B-ALL, T-ALL, AML and plasma cell neoplasms at our lab. I have published extensively in the space of **interpretive nuances of high-sensitivity MRD analysis** including mimics and artefacts in B-ALL MRD (PMID: 34476808) and regenerative immunophenotypic shift in B cell precursors (PMID: 32896101). I co-led the development of a **highly sensitive 15c B-ALL MRD panel** (LLOQ~0.001%) **specifically tailored for patients undergoing CD19-directed therapies** (anti-CD19-CAR-T and blinatumomab, PMID: 37706583).

c) I have **developed pipelines for high-dimensional flowcytometry data analysis** including 5-tube 16 color **deep immune cell profiling** (PMID: 34582592).

3. **Mass cytometry:** During my Mres at Kings College London, I have performed high-dimensional **CyToF** based analysis of immune profiling in patients with Myelodysplastic neoplasm.

4. **Genomics:**

a) I have been actively involved in **standardization and reporting of next-generation sequencing assays** including **targeted DNA and RNA sequencing assays** in acute leukemia and myeloid malignancies. We have developed sequencing assays compatible with Illumina, AVITI and Oxford Nanopore sequencing platforms. We have successfully developed a 137-gene targeted DNA-sequencing panel for acute leukemia and myeloid neoplasm and use it for genomic characterization of patients with hematolymphoid neoplasm as routine practice. I have also helped standardize a novel UMI-based modular targeted RNA-sequencing assay called **NARASIMHA** for detection of chimeric gene fusions (PMID: 32372024).
b) I have contributed to routine implementation of molecular **AML MRD detection using error-corrected sequencing** at our lab**.**
c) Through my WT-DBT India Alliance Intermediate Fellowship, we have developed and validated a **205-gene targeted sequencing panel capable of detecting clinically relevant sequence mutations, translocations and copy number alterations** in patients with Lymphoma. I am currently leading the development of **a simultaneous genetic and epigenetic sequencing assay (Five-letter Seq)** for *ctDNA-based molecular profiling of DLBCL*.
d) Through my LTMT-funded research project, we have standardized a **single-cell multiomic CITE-seq** (whole transcriptome and targeted epitopes) assay for characterizing the cellular/clonal heterogeneity in patients with Acute Leukemia with Ambiguous Lineage.
e) I am well-versed with **conventional molecular diagnostics** routinely performed in molecular hematopathology laboratory such as detection of mutations using allele-specific PCR and fragment length analysis, real time quantitative PCR, chimerism analysis in post-allogenic stem cell transplant patients.
f) I have experience in performing analysis of **Nanostring generated gene expression data** generated using Pancancer Immune Profiling Panel on murine models**.**
g) I have worked extensively in **standardizing and routine reporting of SARS-CoV-2 RT-PCR** at the Hematopathology Laboratory over past 5 years. I have also co-led the effort in analysis and publication of genomic sequencing data of ~1000 SARS-CoV-2 isolates at our institute (PMID: 35685574).

5. **Bioinformatics:** I have been trained in Cancer school of bioinformatics in Kings College London, where I gained familiarity in **basic bioinformatics tools** including basic knowledge in programming languages like R, Python. I have experience in performing high dimensional flow cytometry data analysis using open-source platforms such as R. In our laboratory, we routinely perform custom analysis of sequencing data including variants calling, detection of fusions, copy number alteration analysis and detection of rare events such as MRD using error-corrected sequencing.

6. **Artificial Intelligence:** I have been involved in developing Machine Learning algorithms useful in risk-stratification in patients with hematolymphoid neoplasm. We are currently in the process of developing deep learning tools for ultra-rapid diagnosis and prognostication for hematolymphoid neoplasm using shallow methylation-sequencing or whole-slide imaging.

<u>Key Research Projects at Tata Memorial Centee:</u>

Our project on "Concurrent interrogation of genetic and epigenetic features in Diffuse large B-cell lymphoma and dynamics of recurrence/progression" has been approved by DBT/WT India Alliance.

We have just been funded the ICMR Centre for Advanced Research grant on innovation in Diagnosis, Prognostication, and Monitoring of AML

In addition, I have been actively involved in following projects:

a) **Deciphering the molecular heterogeneity in Acute Leukemia of Ambiguous Lineage at a cellular level (Funded by LTMT)**
b) **Whole Transcriptomic Sequencing (WTS) in pediatric acute leukemia (Funded by Illumina)**
c) Developing and assessing clinical utility of a highly sensitive NGS-MRD assay in pediatric Acute Lymphoblastic Leukemia.
d) Acute myeloid leukemia and the dynamics of relapse.
e) Unravelling the genomic context of relapse initiating cells in adult acute myeloid leukemia.
f) Investigating value of circulating microRNAs and clonal plasma cells in the prediction of therapeutic outcome and prognostication of multiple myeloma.
g) Evaluation of the role of the immune cell profile in the persistence of minimal residual disease and dynamics of the relapse in childhood T-cell lymphoblastic leukemia: a prospective observational study
h) Evaluating the clinical utility of new immunophenotypic markers in the determination of minimal residual disease in childhood acute myeloid leukemia.
i) Investigating the role of flow cytometric immunophenotyping in staging of pediatric solid round cell tumors.
j) Treatment and monitoring of children and adolescents with Chronic Myeloid Leukemia (CML)- A prospective observational study.
k) Indian Childhood Collaborative Leukaemia Group (ICiCLe): A collaborative, multicentre, national trial for newly diagnosed patients with acute lymphoblastic leukaemia.
l) Genomic Landscape of SARS-CoV2 in the city of Mumbai.
m) Immunoprofiling (IP) consortium for COVID-19: Integration of Extended Immune Monitoring (ExImM) and clinical parameters for early prediction of disease trajectory/progression, treatment planning and prophylaxis to improve COVID-19 prognosis.

<u>Teaching experience:</u>

1. I have been regularly involved in **Hematopathology teaching** of Pathology junior residents, Oncopathology DM residents, Hematopathology fellows, Medical Oncology and Pediatric Oncology senior residents. In addition, I have conducted regular workshops on various aspects of high-sensitivity MRD, and molecular techniques relevant to oncology.
2. I have been working as a **mentor and co-guide (Project PI) for PhD student** in Health Sciences at Tata Memorial Centre. I have further served as guide of external projects for MSc students.
3. I have been actively involved as a core member in designing, development and execution of **"Online Teaching Program for Post-Graduates in Pathology (OTPPGP)"**, a collaborative, free-to-register, curriculum-based virtual Pathology learning platform primarily for MD Pathology residents all across the world.
4. I am currently in the process of designing and developing SOP for the proposed **"Tata Memorial Centre Traveling School of Oncopathology (TTSOP)",** an initiative

developed by TMC to improve oncopathology practice and education across the country. The program is scheduled to start from January 2026.

Publications:

- **Total citations =** 884
- **h-index =** 16
- **Google scholar link:** https://scholar.google.com/citations?user=ZHwaj-0AAAAJ&hl=en
- **PubMed link for complete list of 52 publications:** https://pubmed.ncbi.nlm.nih.gov/?term=Chatterjee+Gaurav&sort=date&size=100

List of selected publications:

1. **Chatterjee G**, Dhende P, Raj S, Shetty V, Ghogale S, Deshpande N, Girase K, Patil J, Kalra A, Narula G, Dalvi K, Dhamne C, Moulik NR, Rajpal S, Patkar NV, Banavali S, Gujral S, Subramanian PG, Tembhare PR. 15-color highly sensitive flow cytometry assay for post anti-CD19 targeted therapy (anti-CD19-CAR-T and blinatumomab) measurable residual disease assessment in B-lymphoblastic leukemia/lymphoma: Real-world applicability and challenges. Eur J Haematol. 2024 Jan;112(1):122-136. doi: 10.1111/ejh.14102. Epub 2023 Sep 14. PMID: 37706583.
2. **Chatterjee G**, Sriram H, Ghogale S, Deshpande N, Khanka T, Girase K, Verma S, Arolkar G, Dasgupta N, Narula G, Shetty D, Dhamne C, Moulik NR, Rajpal S, Patkar NV, Banavali S, Gujral S, Subramanian PG, Tembhare PR. Mimics and artefacts of measurable residual disease in a highly sensitive multicolour flow cytometry assay for B-lymphoblastic leukaemia/lymphoma: critical consideration for analysis of measurable residual disease. Br J Haematol. 2022 Jan;196(2):374-379. doi: 10.1111/bjh.17801. Epub 2021 Sep 2. PMID: 34476808.
3. **Chatterjee G**, Dudakia V, Ghogale S, Deshpande N, Girase K, Chaturvedi A, Shetty D, Senger M, Jain H, Bagal B, Bonda A, Punatar S, Gokarn A, Khattry N, Patkar NV, Gujral S, Subramanian PG, Tembhare PR. Expression of CD304/neuropilin-1 in adult b-cell lymphoblastic leukemia/lymphoma and its utility for the measurable residual disease assessment. Int J Lab Hematol. 2021 Jan 12. doi: 10.1111/ijlh.13456. Epub ahead of print. PMID: 33432783.
4. **Chatterjee G**, Sriram H, Ghogale S, Deshpande N, Khanka T, Panda D, Pradhan SN, Girase K, Narula G, Dhamane C, Malik NR, Banavali S, Patkar NV, Gujral S, Subramanian PG, Tembhare PR. Immunophenotypic shift in the B-cell precursors from regenerating bone marrow samples: A critical consideration for measurable residual disease assessment in B-lymphoblastic leukemia. Cytometry B Clin Cytom. 2020 Sep 8. doi: 10.1002/cyto.b.21951. Epub ahead of print. PMID: 32896101.
5. **Chatterjee G**, Pai T, Hardiman T, Avery-Kiejda K, Scott RJ, Spencer J, Pinder SE, Grigoriadis A. Molecular patterns of cancer colonisation in lymph nodes of breast cancer patients. Breast Cancer Res. 2018 Nov 20;20(1):143. doi: 10.1186/s13058-018-1070-3. PMID: 30458865; PMCID: PMC6247766.
6. Shetty A, **Chatterjee G**, Rajpal S, Srivastava T, Gardi N, Mirgh S, Gokarn A, Punatar S, Shetty N, Joshi A, Nair S, Murthy V, Khattry N, Tembhare P, Dikshit R, Chaturvedi P, More A, Kamtalwar S, Chavan P, Bhat V, Patil A, Dhumal S, Bhat P, Subramanian P, Gujral S, Badwe R, Patkar N, Gupta S. Genomic Analysis of AZD1222 (ChAdOx1) Vaccine Breakthrough Infections in the City of Mumbai. Int J Clin Pract. 2022 Feb 11;2022:2449068. doi: 10.1155/2022/2449068. PMID: 35685574
7. Tembhare PR, **Chatterjee G**, Chaturvedi A, Dasgupta N, Khanka T, Verma S, Ghogale SG, Deshpande N, Girase K, Sengar M, Bagal B, Jain H, Shetty D, Rajpal S, Patkar N, Agrawal T, Epari S, Shet T, Subramanian PG, Gujral S. Critical Role of Flow Cytometric

Immunophenotyping in the Diagnosis, Subtyping, and Staging of T-Cell/NK-Cell Non-Hodgkin's Lymphoma in Real-World Practice: A Study of 232 Cases From a Tertiary Cancer Center in India. Front Oncol. 2022 Mar 1;12:779230. doi: 10.3389/fonc.2022.779230. PMID: 35299754.

8. Nathany S, **Chatterjee G**, Ghai S, Moulik NR, Shetty D, Subramanian PG, Tembhare P, Gujral S, Dhamne C, Banavali S, Narula G, Patkar N. Mutational landscape of Juvenile Myelomonocytic Leukemia (JMML)-A real-world context. Int J Lab Hematol. 2021 Dec;43(6):1531-1538. doi: 10.1111/ijlh.13680. Epub 2021 Aug 13. PMID: 34387930.
9. Patkar N, Kakirde C, Shaikh AF, Salve R, Bhanshe P, **Chatterjee G**, Rajpal S, Joshi S, Chaudhary S, Kodgule R, Ghoghale S, Deshpande N, Shetty D, Khizer SH, Jain H, Bagal B, Menon H, Khattry N, Sengar M, Tembhare P, Subramanian P, Gujral S. Clinical impact of panel-based error-corrected next generation sequencing versus flow cytometry to detect measurable residual disease (MRD) in acute myeloid leukemia (AML). Leukemia. 2021 Feb 8. doi: 10.1038/s41375-021-01131-6. Epub ahead of print. PMID: 33558666.
10. Patkar N, Bhanshe P, Rajpal S, Joshi S, Chaudhary S, **Chatterjee G**, Tembhare P, Dhamne C, Prasad M, Moulik NR, Shetty D, Gokarn A, Bonda A, Nayak L, Punatkar S, Bagal B, Sengar M, Narula G, Khattry N, Banavali S, Subramanian PG, Gujral S. NARASIMHA: Novel Assay based on Targeted RNA Sequencing to Identify ChiMeric Gene Fusions in Hematological Malignancies. Blood Cancer J. 2020 May 5;10(5):50. doi: 10.1038/s41408-020-0313-6. PMID: 32372024; PMCID: PMC7200652.
11. Tembhare PR, Sriram H, **Chatterjee G**, Khanka T, Gokarn A, Mirgh S, Rajendra A, Chaturvedi A, Ghogale SG, Deshpande N, Girase K, Dalvi K, Rajpal S, Patkar N, Trivedi B, Joshi A, Murthy V, Shetty N, Nair S, More A, Kamtalwar S, Chavan P, Bhat V, Bhat P, Subramanian PG, Gupta S, Khattry N. Comprehensive immune cell profiling depicts an early immune response associated with severe coronavirus disease 2019 in cancer patients. Immunol Cell Biol. 2022 Jan;100(1):61-73. doi: 10.1111/imcb.12504. Epub 2021 Oct 27. PMID: 34582592.
12. Tembhare PR, Sriram H, Khanka T, **Chatterjee G**, Panda D, Ghogale S, Badrinath Y, Deshpande N, Patkar NV, Narula G, Bagal B, Jain H, Sengar M, Khattry N, Banavali S, Gujral S, Subramanian PG. Flow cytometric evaluation of CD38 expression levels in the newly diagnosed T-cell acute lymphoblastic leukemia and the effect of chemotherapy on its expression in measurable residual disease, refractory disease and relapsed disease: an implication for anti-CD38 immunotherapy. J Immunother Cancer. 2020 May;8(1):e000630. doi: 10.1136/jitc-2020-000630. PMID: 32439800; PMCID: PMC7247386.
13. Tembhare PR, Narula G, Khanka T, Ghogale S, **Chatterjee G**, Patkar NV, Prasad M, Badrinath Y, Deshpande N, Gudapati P, Verma S, Sanyal M, Kunjachan F, Mangang G, Gujral S, Banavali S, Subramanian PG. Post-induction Measurable Residual Disease Using Multicolor Flow Cytometry Is Strongly Predictive of Inferior Clinical Outcome in the Real-Life Management of Childhood T-Cell Acute Lymphoblastic Leukemia: A Study of 256 Patients. Front Oncol. 2020 Apr 24;10:577. doi: 10.3389/fonc.2020.00577. PMID: 32391267; PMCID: PMC7193086.
14. Roy Moulik N, Keerthivasagam S, Pandey A, Agiwale J, Hegde K, **Chatterjee G**, Dhamne C, Prasad M, Chichra A, Srinivasan S, Mohanty P, Jain H, Shetty D, Tembhare P, Patkar N, Narula G, Subramanian PG, Banavali S. Treatment and follow-up of children with chronic myeloid leukaemia in chronic phase (CML-CP) in the tyrosine kinase inhibitor (TKI) era-Two decades of experience from the Tata Memorial Hospital paediatric CML (pCML) cohort. Br J Haematol. 2023 Dec 14. doi: 10.1111/bjh.19251. Epub ahead of print. PMID: 38098201.

15. Patkar N, Shaikh AF, Kakirde C, Nathany S, Ramesh H, Bhanshe P, Joshi S, Chaudhary S, Kannan S, Khizer SH, **Chatterjee G**, Tembhare P, Shetty D, Gokarn A, Punatkar S, Bonda A, Nayak L, Jain H, Khattry N, Bagal B, Sengar M, Gujral S, Subramanian P. A novel machine-learning-derived genetic score correlates with measurable residual disease and is highly predictive of outcome in acute myeloid leukemia with mutated NPM1. Blood Cancer J. 2019 Oct 1;9(10):79. doi: 10.1038/s41408-019-0244-2. PMID: 31575857; PMCID: PMC6773777.
16. Tembhare PR, Subramanian Pg PG, Ghogale S, **Chatterjee G**, Patkar NV, Gupta A, Shukla R, Badrinath Y, Deshpande N, Narula G, Rodrigues P, Girase K, Dhaliwal D, Prasad M, Shetty D, Banavali S, Gujral S. A High-Sensitivity 10-Color Flow Cytometric Minimal Residual Disease Assay in B-Lymphoblastic Leukemia/Lymphoma Can Easily Achieve the Sensitivity of 2-in-10(6) and Is Superior to Standard Minimal Residual Disease Assay: A Study of 622 Patients. Cytometry B Clin Cytom. 2019 Jun 14. doi: 10.1002/cyto.b.21831. [Epub ahead of print] PubMed PMID:31197916.
17. Patkar N, Kakirde C, Bhanshe P, Joshi S, Chaudhary S, Badrinath Y, Ghoghale S, Deshpande N, Kadechkar S, **Chatterjee G**, Kannan S, Shetty D, Gokarn A, Punatkar S, Bonda A, Nayak L, Jain H, Bagal B, Menon H, Sengar M, Khizer SH, Khattry N, Tembhare P, Gujral S and Subramanian P (2019) Utility of Immunophenotypic Measurable Residual Disease in Adult Acute Myeloid Leukemia—Real-World Context. *Front. Oncol*. 9:450. doi: 10.3389/fonc.2019.00450
18. Rajendra A, Jain H, Bonda VNA, Nayak L, Tembhare P, Shetty D, Thorat J, Jain H, Subramanian PG, Patkar N, **Chatterjee G**, Khattry N, Gokarn A, Punatar S, Mokal S, Bagal B, Sengar M. Outcomes and prognostic factors in adolescents and young adults with ALL treated with a modified BFM-90 protocol. Blood Adv. 2021 Mar 9;5(5):1178-1193. doi: 10.1182/bloodadvances.2020003526. PMID: 33635331.
19. **Chatterjee G**., Gujral S., Subramanian P.G, Tembhare P. Clinical relevance of multicolour flowcytometry in Plasma Cell Disorders. Indian J Hematol Blood Transfus (2017). doi:10.1007/s12288-017-0822-z
20. Tembhare PR, Ghogale S, Ghatwai N, Badrinath Y, Kunder N, Patkar NV, Bibi AR, **Chatterjee G**, Arora B, Narula G, Banawali S, Deshpande N, Amare P, Gujral S, and Subramanian PG. Evaluation of New Markers for Minimal Residual Disease Monitoring in B-Cell Precursor Acute Lymphoblastic Leukemia: CD73 and CD86 Are the Most Relevant New Markers to Increase the Efficacy of MRD Assay. Cytometry Part B 2016
21. **Chatterjee G**, DasGupta S, Mukherjee G, Sengupta M, Roy P, Arun I, Datta C, Mishra PK, Banerjee S, Chatterjee U. Usefulness of Wieneke criteria in assessing morphologic characteristics of adrenocortical tumors in children. Pediatr Surg Int. 2015 Jun;31(6):563-71

Notable Abstracts / Platform Presentations:

Over the years, I have actively contributed to national and international scientific forums through abstracts, posters, workshops, and invited faculty roles. Selected highlights include:

1. Poster on “Clinical relevance of peripheral blood measurable residual disease assessment in newly diagnosed multiple myeloma after initial therapy: a prospective study” at ICML, Lugano 2025.
2. Participated as a Delegate in London Calling 2025 organized by Oxford Nanopore.

3. e-poster on "Myelodysplasia-Related Mutations in Adults and End-of-Induction Measurable Residual Disease Are Associated with Inferior Outcome in Patients with Acute Leukemia of Ambiguous Lineage" at ASH 2024.
4. Presented poster on "A series of pediatric myeloid neoplasms harboring mutations in genes associated with germline predisposition in myeloid malignancies" at VEPTC 2023, Barcelona.
5. Presented poster on "Online Teaching Program for Post Graduates in Pathology: A Virtual, Collaborative, Free-to-Register, Curriculum-Based, Long-Term Pathology Learning Portal" at USCAP 2021.
6. Presented psoter on "Genomic Landscape of Juvenile Myelomonocytic Leukemia: A Real World Context" at ASH 2019.
7. Poster on "A Cost-Effective, High Sensitivity 10-Color Single Tube Flowcytometry Based B-Cell Precursor Acute Lymphoblastic Leukemia Minimal Residual Disease (MRD) Assay With Study Of Artifacts And Mimics" at EHA 2017.
8. Platform poster presentation on "Utility of the New Versus Old Immunophenotypic Markers in the Flow Cytometric Immunophenotyping of Multiple Myeloma" at the 16th International IMWG Myeloma Workshop 2017.
9. Oral Presentation on "Development of single-tube 15 colour lymphoma screening tube" at ESCCA 2016.
10. I have worked as a core organizing member of multiple meetings/workshops at ACTREC, TMC including: (a) 16th TCS 2024; (b) 1st Genomics Workshop on NGS & Bioinformatics 2024; and (c) 1st Indian B-ALL MRD Consensus Recommendation Meeting & Workshop 2025.

Guidelines and other extramural responsibilities:

1. I was an integral part in the Indian **B-ALL FCM MRD consensus guidelines developing workshop** organized by our department at ACTREC, Tata Memorial Centre. As part of this venture, I have led a **multicentric effort to standardize assay sensitivity thresholds (limit of detection and lower limit of quantitation)**. These guidelines are due to publish in 2026.
2. I have also contributed to the **NCG Guideline Development Group for Adult Hematolymphoid Cancers**, 2019 and 2025-26.
3. I have been nominated as the **Innovation Ambassador of Tata Memorial Centre for HIIC (HBNI- Institution Innovation Council)** for 2024-25.
4. I have been working as **Reviewer** of the BJH, IJHBT, PHO, SAJC, JCP, Clinical Cytometry and Frontiers in Oncology.

Other Honors and awards:

- Awarded **"The Best Published Paper Annual Award 2021 (Clinical)"** for the paper titled "Immunophenotypic shift in the B-cell precursors from regenerating bone marrow samples: A critical consideration for measurable residual disease assessment in B-lymphoblastic leukemia. Clinical Cytometry 2021; 100:434–445. DOI: 10.1002/ctob21951" at the 13th Annual Conference-cum-Workshop organised The Cytometry Society of India, 2021
- Secured Honors marks in Pharmacology in M.B.B.S.
- Awarded 1st place in College, State & Zonal level and 3rd/ 4th place in National level in ISHBT National Hematology Quiz competition, 2013& 2014.

- Awarded best poster at 8th TCS & 16th Indo-US Cytometry workshop, TMH, Mumbai, 2015
- Awarded best oral paper at Hematocon, 2016, India.
- Awarded 2nd best oral paper at State-of-the-art myeloma meet, PGIMER, Chandigarh, 2016
- Awarded best oral paper at NIIH Platinum jubilee conference, Mumbai, 2017
- Awarded 2nd best oral paper at 18th Indo-US Cytometry workshop, SGPGI, Lucknow, 2017
- Awarded JC Patel best paper at 40th annual conference of Mumbai Hematology Group, 2017

Membership of Professional Organisations:

- USCAP
- The Cytometry Society, India
- Molecular Pathology Association of India

References:

1. Prof Sumeet Gujral Professor, Department of Pathology, 8th floor, Annexe Building, Tata Memorial Centre, Mumbai-400012 **Email: flowtmh@gmail.com** **Mobile:** +91 9820523962	**2. Dr Anita Grigoriadis** Senior Lecturer in the School of Cancer and Pharmaceutical Sciences at King's College London (KCL), the School Lead (International) and Training Lead for the CRUK KHP Centre, UK **Email: anita.grigoriadis@kcl.ac.uk**	**3. Prof Ghulam Mufti** Head of the department of haematological medicine at Guy's, King's and St Thomas's hospitals, King's College London, UK **Email: ghulam.mufti@kcl.ac.uk**

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Date: 28th October 2025